

## The role of *Typha* sp. in affecting the radium behavior in the water column of a salt coastal lagoon

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**Abstract.** The radium accumulation and releasing in *Typha Domingenes* Pers. foliage in a coastal lagoon was studied by in situ and lab experiments. From leaf sample analysis and lab experiments, adsorption experiments followed by procedure sequential extraction, it was observed the importance of the ionic exchange for the foliar accumulation. The higher the salinity, the lower the accumulation in leaves. Decomposition of leaves was studied by litterbag methods during a period of 6 months. At the end of the experiments the major cation losses were 99% of K, 76% of Na and 51% of Mg. On the other hand the amount of Ca in the residue increased seven times as well as increased the total activity of <sup>226</sup>Ra and of <sup>228</sup>Ra (seven and at least three times, respectively). In agreement with this observation, laboratory experiments showed that the material was able to adsorb quite a 100% of the added Ra and its adsorption capacity was estimated at 5 meq (Ba<sup>2+</sup>)/g<sub>detritus</sub>. Radionuclide releasing by sequential extraction of the detritus material followed a little bit different dynamic from the green leaves, showing that important fraction of the radium can be retained in the litter by humidified material.

### 1. INTRODUCTION

The Buena lagoon is a shallow salt lagoon whose water column contains abnormal high concentrations of radium isotopes. The Ra-226 concentration ranges from 0.05 to 0.89 Bq/L, while the Ra-228 concentration ranges from 0.07 to 2.01 Bq/L. The salinity varies between 9 and 42 ‰ and the pH between 4.2 and 7.3. The radionuclide concentrations decrease in seawater direction, while the pH and its salinity increases in the same direction [1]. The lagoon and its wetland are largely covered by aquatic vegetation, mainly *Typha Domingensis* Pers. *Typha* sp. (cattails) is a rooted aquatic macrophyte plant, commonly found in tropical and temperate wetlands, whose behaviour has been studied with the aim to use it in polluted systems and for energy production [2, 3, 4].

Rooted macrophyte plants can accumulated minerals from water column and sediment, being the free accessibility of water and solutes an important factor for foliage uptake to cell wall. However, there is not a consensus yet on which is the main route for nutrients uptake by plant, if foliar or root uptake [5, 6]. From literature data gathering the average value of concentration ratio, CR, for radium from water to plant wet foliage was estimated at 55 [7]. Iyengar and Rao reported values of radium CR for different rooted macrophytes ranging from 30 to 1400, being the recommended value 100, while the CR from sediment to wet aquatic plant foliage was estimated at 0.014, independently of the water contribution [8].

Plant decomposition is a process that involves physical, chemical and biological events resulting in the production of CO<sub>2</sub>, minerals (as example calcite: CaCO<sub>3</sub>), water and humified organic matter. Basically, the decomposition occurs in three steps. The first step of process involves physical leaching of nutrients and dissolved organic compounds (soluble carbohydrates, lipids and polyphenols), which account for 6 to 40% dry weight loss within 15 days. The leaching phase is followed by microbial decomposition by bacteria and fungi, leading to decomposition of structural compounds such as cellulose, hemicelluloses and lignin. Then, a third step characterized by the reduction of bacterial activity takes place until the decomposition stops, remaining the refractory foliage material [9,10, 11].

*Typha domingensis* Pers. is the unique *Typha* sp. specie found in Brazil, it can be found in most wetlands and coastal lagoons. Its foliage is responsible for 60 to 70% of the whole plant biomass and for the larger nutrient deposit of the plant [2]. Its organic matter content is higher than 85% and the wall cell comprises around 70% of the foliage [12]. The foliage is composed by circa of 20% of polyphenols and 40% of soluble carbohydrates [13]. Furtado, comparing the constituents in green leaves and in leaf detritus observed that the detritus presents 31% less of organic matter, 50% less of cell wall, 62% less of soluble carbohydrate, 21% less of lipids and 80% less of the tannin than the green leaves [12]. After six month of *Typha* foliage decomposition experiment, Couto observed a loss of 40 to 80% of potassium, 40 to 90% of sodium, 60 to 80% of magnesium, 40 % of calcium and iron and 40 to 60% of manganese [14].

The evaluation of the role of *Typha* in the Buena lagoon system is important to assess the radium cycling and its reflection on the observed high radium concentration on lagoon water column. On the

other hand, the obtained information can be useful regarding the understanding of radium behaviour in natural aquatic systems as well as the use of *Typha* sp. for recovering radium from aqueous effluents of uranium mining and milling. The aiming of this research was to study the role of *Typha* sp. in the cycling of radium in the lagoon considering its utilization for remediation of mine drainage wastes.

## 1.1. Material and Methods

### 1.1.1 The study area

The Buena Lagoon belongs to the São Francisco de Itabapoana county in the Rio de Janeiro State, Brazil, between the coordinates 21° 23' - 21° 24' S and 41° 00' - 41° 03' W. Its watercourse surface area is 0.11 km<sup>2</sup> (113,000 m<sup>2</sup>), whose length is 5.4 km and the average width is 21 m. Its wetland has an area of 670,000 m<sup>2</sup>, being 518 000 m<sup>2</sup> completely covered by *Typha Domingensis* Pers., and its catchments area comprises 7.5 km<sup>2</sup>. The lagoon is shallow (maximum depth 3 m) and it is separated from the ocean by a barrier of sand, which width is approximately 15 meters and eventually, direct seawater inflow can occur.

### 1.1.2 Sampling and Chemical Analysis

The lagoon map was digitalized and information about size, width, catchments and wetland areas were obtained by IDRISIS software. Neither sampling station 1 nor sampling station 7 were found cattails, showing that due to their characteristic water, at station 1, low pH 4, (at the lagoon head) and at station 7 (located close to sea) high salinity 42‰, the plant growing was not possible. At the other stations, plant population was determined by counting the number of plants per square meters. A 1m x 1m square was randomly placed at five points in each sampling point. All the cattails within the square were counted. From one of square the total leaves were sampled. These samples were collected in two periods: one of dry and the other of rain seasons. At the laboratory the samples were cleaned with distilled water, air-dried, weighted and dried in a forced air oven at 80°C. After that the samples were burnt at 450°C and 10 g of ashes were digested with 3:1 mixture of nitric and chloride acid. <sup>228</sup>Ra and <sup>226</sup>Ra were analysed by total alpha and beta counting [15]. The accuracy of such determination is routinely tested by inter-laboratory exercises organized by the EPA/USA and the Radionuclide Metrology Division of IRD/CNEN [16].

Sediment and water samples were collected from the same points of cattail sampling. The sediment was collected with an auger to a depth of 10 cm and transferred to plastic bags. Immediately after the sampling, sediment pH and Eh measurements were performed in the wet sediment. At the lab the sediments were air dried and passed through a 2 mm sieve. After 20 days, the samples were analysed by gamma spectrometry Ra-226 (Pb-214, 614 keV) and Ra-228 (Ac-228, 911 keV).

About 3.0 L of water in a polyethylene bottle were collected at each sampling point, in two different seasons – a rainy and a dry. Eh, pH and conductivity were measured directly at the site. A Millipore membrane filtered the water and the sample was acidified with nitric acid supra pure for the determination of the radionuclides and cations. Major cations were analysed by inductively coupled plasma-mass spectrometry, ICP-MS, Perkin-Elmer ELAN 5000, after a dilution 1:1000 with water, Milli-Q, applying the so-called “TotalQuant” method and In and Tl as internal standards. A total of 27 elements were used for the equipment mass efficiency calibration. Using 1-liter sample, <sup>228</sup>Ra and <sup>226</sup>Ra were analysed by total alpha and beta counting [15].

### 1.1.3 In situ decomposition

The *Typha* leaves were cut into 15 cm segments and oven dried at 80°C to constant weight and 10 g of dry leaves were placed in nylon litter bags (1mm mesh size). Forty 20 X 15 cm size bags were taken in total. The bags were placed on the bottom of the lagoon, submerged under 30-50 cm water, at sampling station 5. Five bags were taken out at different intervals of time and then they were washed in lagoon water and rinsed in distilled water. The plant material was dried in hot air oven at 80°C, weighed, and powdered in quartz mortar for chemical analysis of Na, K, Ca, Mg, Ra-228 and Ra-226. The radium concentration in the six months material could not be determined, due to the small quantity of recovered material.

### 1.1.4 Laboratory adsorption experiments

Green leaves: 15 g of green leaves in pieces of 10 cm were placed in 250 ml Teflon centrifuge tube and immersed in 50 ml of a solution, pH 6, containing 32 Bq of Ra-228 and 1g/L NaCl, for 8 days. Then the

leaves were taken out of solution, washed with distilled water, fractionated into little pieces and placed in polyethylene bottles for counting on a germanium detector.

Detritus: the six-month foliage detritus were ground in a quartz mortar to pass a 2 mm screen. Then, 0.05 g of the samples was placed in a Teflon tube and 50 ml of lagoon water and 12 Bq of Ra-228 were added to the detritus. After 24 hours the detritus was filtered in a Millipore membrane 5 cm diameter, 0.45  $\mu\text{m}$ . The filter was washed with distilled water and analysed by a germanium detector.

The adsorption capacity of the detritus was evaluated by adding 0.05 g of the detritus material to 50 ml solution, pH=6.8, containing 60 mg of  $\text{Ba}^{++}$  and 23 Bq of Ba-133. The sample was allowed to stand for 24 hours before filtering in a Millipore membrane 5 cm diameter, 0.45  $\mu\text{m}$ . The filter supporting the detritus was washed with distilled water and analysed on a germanium detector

## 2. RESULTS

The radium concentration in foliage was inversely correlated with the logarithm of addition of Ca and Mg concentrations in the water ( $r=0.73$  for Ra-226 and  $0.55$  for Ra-228,  $P<0.01$ ), table 1, supporting the hypotheses that Ra was accumulated and distributed within the plant by the same mechanisms as the divalent cations and that a probable competition among the cations for adsorption sites in the plant is occurring [7]. It was not observed correlation between the pH in the water or in sediment with the radium concentration in foliage.

Amounts of metals removed from water by cattail depend upon cattail biomass and concentration of metals in plants. Foliage biomass ranged from 230 to 1180 g dry matter. $\text{m}^2$ . As it was not found statistical significant difference of the plant biomass between the sampling seasons and among the sampling stations, the average biomass could be estimated as 610 g dry matter. $\text{m}^2$ . The values of biomass fall into the observed range for *Typha* in other Brazilian coastal lagoons (105 to 1600 g dry matter. $\text{m}^2$ ) [13,14]. The relationship between wet to dry mass and wet to ash mass point out to 72% of water and 2.3% of ash in the foliage. Between the sampling station 2 and the sampling station 5, the wetland area is approximately 518,000  $\text{m}^2$ , resulting in a foliage total dry mass of 316 ton (wet mass 1129 ton and ash mass 24 ton) in the lagoon. The Ra concentration in the foliage was correlated with the Ra concentration in the water column Ra-228 ( $r=0.541$ ,  $P<0.05$ ) and Ra-226 ( $r=0.499$ ,  $P<0.05$ ). However no correlation was found among the Radium concentration in sediment and in foliage. As it is expected, no statistic difference was found between the concentration ratios CR from water to plant wet foliage of the two isotopes. The concentration ratio CR ranged from 6 to 125 and the geometric average value was estimated as 33. This CR average value is lower than that one observed in rooted macrophyte, showing that *Typha* in that lagoon system is not specially a good radium accumulator.

By testing the difference among geometric average (two tailed t-test) and comparison of geometric standard deviations (F-test), the station 2 was identified as the station of more elevated concentration of radium isotopes. As it was not found significant statistically difference among the radium concentration in the other stations, the geometric average concentrations of Ra-226 and Ra-228 in wet foliage was determined for station 2: 11.7 and 21 Bq/kg respectively, while for the other stations (3,4 and 5): 5.3 Bq/kg for Ra-226 and 7.3 Bq/kg for Ra-228. In the gross, it is possible to estimate the total activity of radium in the cattail foliage. By IDRISIS software, the wetland area at station 2 was estimated as app. 64,000  $\text{m}^2$ , corresponding to a wet foliage mass of 139 ton and 2 MBq of Ra-226 and 3 MBq of Ra-228. The other 990 ton of foliage would contain 5 MBq of Ra-226 and 7 MBq of Ra-228, bringing up a total of 7 MBq of Ra-226 and 10 MBq of Ra-228 in 300 ton of dry foliage (24 tons of ashes). As the activity of radium isotopes in water column of Buena lagoon were estimated as 20 MBq of Ra-226 and 40 MBq of Ra-228 [1], the foliage would contain app. 20 % of total radium of the system water-foliage. But in the case of foliage cutting aiming decontamination, it is interesting to notice that this amount of radium isotopes would be contained in a waste of 24-ton ashes.

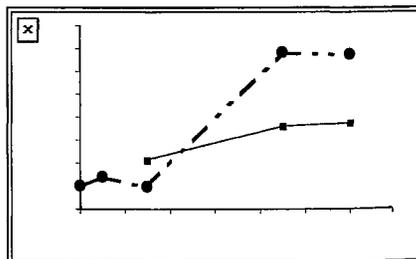
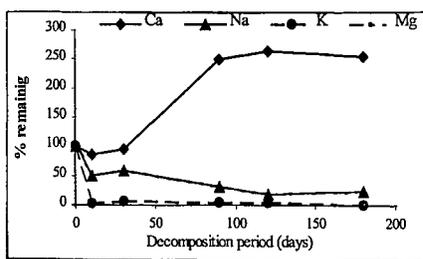
**Table 1.** Concentration values of Ra-226, Ra-228 and magnesium plus calcium in the different compartments of the lagoon.

Sampling Station	Foliage		Biomass g <sub>dry</sub> . m <sup>-2</sup>	Sediment		Water			pH	Cond. mS
	Ra-226 Bq/kg <sub>fresh</sub>	Ra-228		Ra-226 Bq/kg	Ra-228	Ra-226 Bq/L	Ra-228 Bq/L	Mg+Ca mg/L		
2	3.9	20	340	14	43	0.23	0.93	333	6.1	10.0
2	17	28	530	34	94	0.28	0.81	51	6.1	2.8
2	21	31	660	39	82	0.28	0.66	63	n.a	3.5
2	14	11	630	28	90	0.25	0.56	49	5.9	2.5
3	1.3	3.7	580	34	89	0.20	0.36	100	6.6	5.4
3	9	12	420	30	79	0.23	0.50	66	6.2	4.3
3	15	7.4	300	36	80	0.18	0.46	90	n.a.	4.6
3	4.2	4.6	740	31	76	na	na	162	4.4	6.9
4	4.6	15	450	180	233	0.21	0.30	115	6.7	5.7
4	12	30	230	113	491	0.17	0.22	82	6.2	3.8
4	8.2	3.5	760	75	301	0.14	0.31	107	n.a.	5.3
4	0.61	2.8	1180	83	350	0.12	0.15	107	6.4	3.0
5	10	17	540	na	na	0.21	0.39	135	6.8	5.2
5	13	3.8	970	na	na	0.11	0.19	178	6.4	6.3
5	3.2	7.9	670	na	na	0.08	0.08	101	n.a.	3.9

n.a. Not analysed.

**2.1. In situ decomposition**

The data from detritus decomposition show that 65% of dry weight was lost within 6 months. The value of decomposition rate calculated by the decay exponential function [10] was high, 0.0072 d<sup>-1</sup>, which means that the remaining material after one year would be equal to 7% of the initial material. During decomposition potassium was lost quickly, 98% of the total content during the first 15 days. The changes in Na and Mg followed a similar pattern, losing ca. 50% of their initial amount during the first step of decomposition, after that the loss was quite slower. About 20% of sodium and magnesium was still left after 6 months



**Figures 1 and 2:** Changes in K, Na, Mg, Ca and Ra content of decomposition plant material.

Completely different was the calcium behaviour during the decomposition. The calcium content remained almost constant during the first month; after that, increased reaching values 2.6 times more elevated than the initial one (figure 1). The behaviour of radium was quite similar to the calcium one (figure 2) and a significant correlation were found between their contents in detritus ( $r=0.89$ ,  $P=0.05$ ), suggesting similar processes to both cations sorption.

According to the decomposition phases, the quick loss of K, Na and Mg in the beginning is related to the leaching of these nutrients, which are weakly binding to the plant tissue. The calcium and radium behaviour during the first 30 days could be justified by its accumulation as insoluble deposits in plants [7], remaining incorporated to the material. After that, however, their content increasing could be attributed either to contamination by silt or molluscs or to their sorption on detritus sites. In this case, decomposition of structural compounds by microbial process, which would breakdown large molecules in smaller ones, would expose smaller molecules with higher sorption capacity than the original one.

## 2.2. Laboratory adsorption experiments

The adsorption experiments showed that 69 % (22 Bq) of initial radium was retained on the green leaves, pointing out to a passive process of adsorption on cell walls.

The sorption of radium by detritus was almost 100% of the added radium (11.9 Bq), showing that the detritus can really adsorb radium and that the observed increase of the radium and calcium concentrations in decomposition field experiment could be attributed to sorption process, instead of contamination. The sorption capacity experiment resulted in sorption of 28% of Ba-133, corresponding to 17 mg de Ba<sup>++</sup> accumulated on 0.05 g of detritus, meaning a sorption capacity of 340 mg of Ba/g<sub>detritus</sub>.

The contaminated detritus and green leaf were exposed to 0.01 N of ethylenediaminetetraacetic acid (EDTA) solution during 1 minute, and then the solution was analysed by gamma spectrometry. The activity concentration in the solution was less than minimum detectable concentration, pointing out to less than 6% of the radium and barium accumulated by the materials could be recovered by EDTA, so the radium binding with the material are not so weak as observed in algae foliage whose experiment of chemical desorption with EDTA resulted in a high recovery of radium [5]. Then a methodology frequently used to sediments was applied, taking into account that the chemical behaviour and ion competitions should be similar in both cases. So, the plant materials were submitted to a sequential extraction procedure using MgCl<sub>2</sub> solution, 1M, pH 7, which due to its chemical similarity and high cation concentration can compete with radium for the exchange sites on material. Extraction with 0.6 M HOAc/ 1 M NaOAc (pH=5), which could release radium from adsorption sites, specific adsorption, and radium taking part in carbonate compounds and oxidation of organic matter with 30% H<sub>2</sub>O<sub>2</sub>- 0.02 M HNO<sub>3</sub>, 80°C, to release that radium strongly bound to organic fraction and sulphite [17].

Approximately 60 % of radium were recovered by magnesium chloride and acetic acid leaching, pointing out the importance of ionic exchange to the radium adsorption on both materials and denoting that adsorbed radium from detritus and foliage can be released to water column in case sea water inflows in the lagoon or during a dry weather long period, increasing the lagoon water ionic strength. It is interesting to notice that even though oxidation of organic matter was performed around 20% of radium remained firmly bound on the green foliage fibers and therefore would not be released under natural conditions. Only at the end of leaf decomposition this fraction could return to the water column, unless radium is associated with refractory plant material buried in the sediment [4].

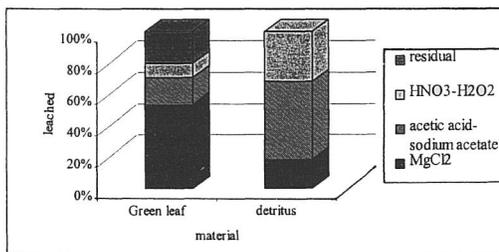


Figure 3: Radium speciation in plant materials

## 3. CONCLUSION

*Typha* foliage can adsorb radium by passive binding to cell walls. The accumulation by foliage is a dynamic process since a significant fraction can be released by increasing water salinity. However, a radium fraction will be so firmly bound to the plant tissue that it will hardly be released from foliage, no matter the changes in water conditions. Detritus can efficiently adsorb radium; however, as for green leaves, changing in the

water conditions could also promote the return of radium to water. Joining to sediment, the detritus will increase the sediment sorption capacity. It is possible that radium remains fixed to the humified material joined to sediment for a period of time, but at the end of the organic matter decomposition it could return to water column, then the detritus serves at least as a temporary storage compartment. The use of *Typha* foliage for radium remediation of water and mine effluents is depreciated by the amount of generated waste. Nevertheless, *Typha* detritus traps radium from aqueous solutions efficiently and its utilization for mine drainage waste remediation should be investigated.

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### References

- [1] Lauria D.C. and Godoy J.M., Journal of Environmental Radioactivity, accepted for publication.
- [2] Garver E.G., Dubbe D.R. and Pratt D.C., Aquatic Botany **32** (1988) 115-127.
- [3] Dunbanbin J.S., Pokorny J., Bowmer K.H., Aquatic Botany **29** (1988) 303-317.
- [4] Dunbanbin J.S., Bowmer K.H., The Science of Total Environment **11** (1992) 151-168.
- [5] Kalin M. and Sharma H.D., (In International Atomic Energy Agency Technical Document (IAEA-SM-275/10,1981)) pp. 247-262.
- [6] L.J.Jackson, The Science of Total Environment (1998) 219- 223.
- [7] Williams A.R. , (Technical Report Series 310, International Atomic Energy, Vienna, Vol.1, 1990) pp.487-508.
- [8] Iyengar M.A.R. and Rao K.N., (The Environmental Behaviour of Radium, Technical Report Series 310, International Atomic Energy, Vienna, Vol.1, 1990) pp.59-128.
- [9] Shama K.P. and Gopal B., (*Wetland Ecology and Management*, National Institute of Ecology and International Scientific Public, 1982) pp. 321-334.
- [10] Kulshreshtha M. and Gopal B., (*Wetland Ecology and Management*, National Institute of Ecology, and International Scientific Public, 1982) pp. 279-292.
- [11] Howard-Williams C. and Howard-Williams W., Aquatic Botany **4** (1978) 257-267.
- [12] Furtado A., (Master Thesis Degree, Departamento de Ecologia, UFRJ, Rio de Janeiro, 1992) 106 p.
- [13] Júnior M.N.A., (Master Thesis Degree, Departamento de Geoquímica, UFF, Rio de Janeiro, 1991) 106p.
- [14] Couto E.C.G., (Master Thesis Degree, Departamento de Geoquímica, UFF, Rio de Janeiro, 1989) 242p.
- [15] Godoy J.M., D.C. Lauria, M.L.D.P. Godoy, R.P. Cunha, R.P., Journal of Radioanalytical and Nuclear Chemistry Articles, **182** 1 (1994), 165-171.
- [16] Vianna M.E.C.M., Tauata L., Oliveira A.E., Oliveira J.P., Clain A.F. and Ferreira A.C.M. Appl.Radiat.Isto. **49** (1998) 1463-1466.
- [17] W.Salomons, U.Förtner, in *Metals in Hydrocycle*, Springer-Verlag, Berlin, 332 (1984).